

**Product Name:** Oligo cleaning-up kit  
**Catalog no.:** EZC202  
**Size:** 50 preps

**Related products:**  
 Mini Spin columns w/collection tubes  
 Tini Spin columns w/collection tubes  
 DNA (ChIP) clean-up and concentration kit (Tini column)

**Catalog No.**  
 EZC101  
 EZC106  
 602

### Description:

This kit is designed for general cleanup of oligonucleotides and DNA up to 10kb from enzymatic reactions (e.g., labeling, dephosphorylation, restriction, and tailing) with silica base Mini Spin Column or Tini spin column. The recover range for oligo and DNA is from 17bp to 10kb. Tini spin column can also be used for concentrating DNA sample by using the same protocol but cut down all the solutions to 1/2 to 1/3. The elution volume is as low as 5µl. The catalog number for cleanup kit with Tini column is Cat#603.

### Feature:

- Easy and rapid with 10 min. procedure using spin column.
- High DNA recovery (17bp-10kb)
- Cleanup oligonucleotides and DNA from mini prep and enzymatic reactions (eg. labeling, restriction, and dephosphorlation....)
- Remove Salt, primers, enzymes, dNTPs, and other impurities

### Kit Contents:

Components	Oligo Cleaning up kit 50 Preps (cat# EZC202)
Oligo Cleanup Binding Buffer (concentrated) *	12.5 ml
5xWash Buffer**	12 ml
Mini spin column with collection tube***	50

\*Add 100% isopropanol before use: add 19 ml 100% isopropanol to 12.5 ml Oligo Cleanup Binding Buffer.

\*\*Add 100% ethanol before use: add 48ml 100% ethanol to 12ml 5xWash Buffer.

\*\*\*Mini Spin columns can be order separately for leftover solutions (cat#EZC101, \$39 for 100 columns)

### Caution:

Oligo Cleanup Binding Buffer contains chaotropic salt. Please use proper safety precautions and always wear gloves when handling the reagent. Avoid contact with skin, eyes or clothing. In case of accidental spill or contact, wash thoroughly with water, seek medical advice if necessary.

### Procedures:

1. Mix 10 volumes of the **Oligo Cleanup Binding Buffer** with 1 volume of reaction sample. For example, add 500µl binding buffer to a 50µl reaction sample. For DNA fragments >100bp, only 5 volumes of binding buffer are required.
2. Load the sample mixture on the **Mini Spin Column** and spin in a microcentrifuge for 1 min at full speed (about 10,000 rpm). Do not load more than 700 µl of sample on Mini spin column (or 350µl on Tini spin column) at one time. Discard the flow through and load more sample mixture if needed.
3. Wash the column by adding 750 µl of **1x Wash Buffer (Ethanol added)** and centrifuge for 1 min.
4. Wash once with 500 µl of 80% ethanol and centrifuge for 1 min (optional).
5. Discard flow through and place the column back in the same tube. Cut off the cap on the column (this will help to remove ethanol completely) and spin for 2 min. **Note:** this step is important since the residual ethanol may affect downstream applications)
6. Place the column in a clean 1.5 ml micro-centrifuge tube.
7. Add 30 µl or more **TE** (10mM Tris,Cl, 1mM EDTA, pH8.0) or **ddH<sub>2</sub>O** (preheated at 65°C for better yield) to the **center** of the column and leave at room temperature for 5 min. Spin the column for 1 min to elute the DNA from the column. For Tini spin column, add as low as 5µl **TE** or **ddH<sub>2</sub>O** for elution.